

Virtual IARPA TEI-REX Proposers' Day

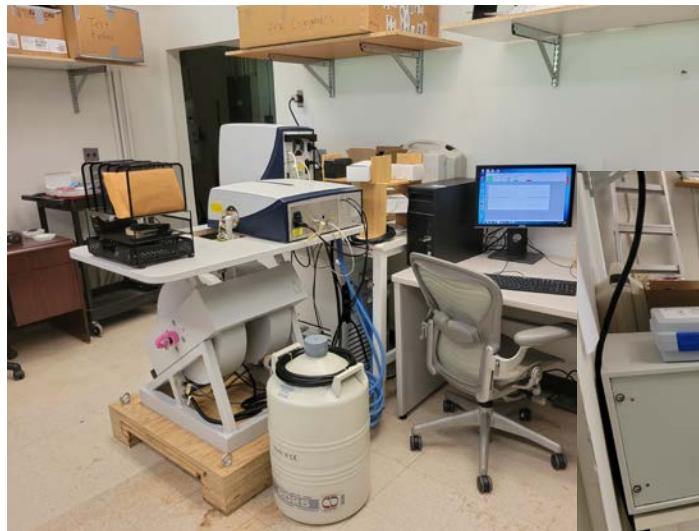
Dr. Robert Bruce Hayes, CHP, PE
Associate Professor
Nuclear Engineering Department
North Carolina State University

[Radiological Detection and Nuclear Assay](#)

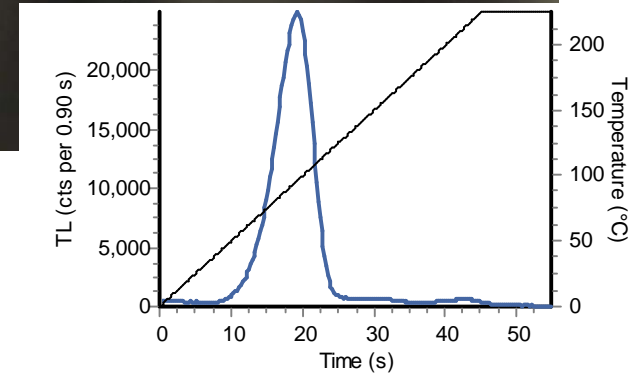
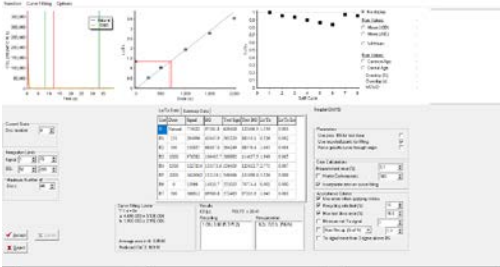
Wed, Sept 29

Electron Paramagnetic Resonance (EPR)

- A free radical is simply an unpaired electron
- Identical physics to that of NMR but EPR sees free radicals rather than free protons
- EPR is a nondestructive means to measure free radicals in a solid.



TL/OSL



EPR biosimetry examples

- **Antlers** which unlike horns are annually shed by fauna allowing regional biosimetry
- **Walrus tusks** which can give results on underwater nuclear activities with which they have contacted or other arctic conditions
- **Shells, mollusks and snails** can provide regional historical radiological conditions
- **Plastic materials** can be used but generally have low sensitivity and short (days to weeks) half lives and vary from one material to another

Thermal and Optically Stimulated Luminescence Biodosimetry Examples

- All insulator materials on a person which have a crystalline structure
 - This does require samples to be largely **translucent**
- Silica type materials including natural quartz, quartzite, feldspars and zircons
 - **Dirt** from money including both paper and coin
 - This requires silica components which have been light shielded
 - **Natural rocks**
 - This requires the accumulated natural background is small or comparable to the dose of interest

Solid State Biodosimetry

based on published research

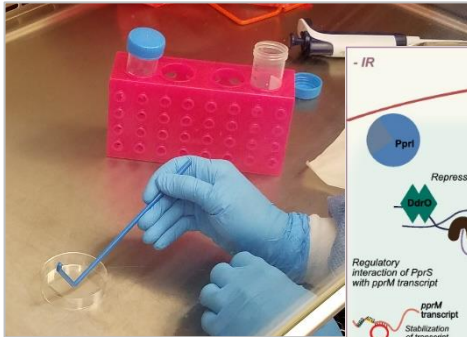
EPR

- Tooth enamel biopsy
 - Requires dentist rather than phlebotomist/nurses
- Confectionary
 - Requires edibles
- Fingernails
 - Low sensitivity and transients
- Buttons
 - Not well characterized

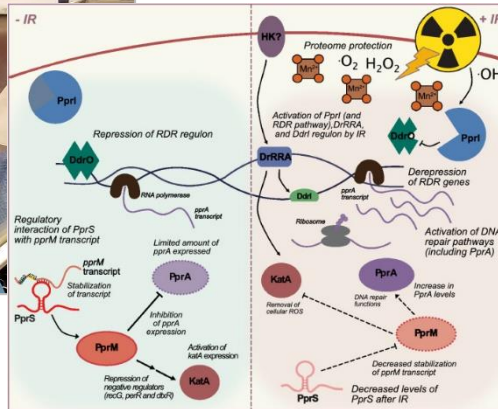
TL/OSL

- Watch PC board
 - Destructive unless reassembling watch
 - Sample preparation requires further research
- Surface mount resistors in personal electronics
 - High sensitivity but requires sufficient electronic components and is in principle destructive





In vitro modeling of skin microbiome



In vivo characterization of radiation response (Villa et al., 2021)

Project Overview

- Our approach focuses primarily on genomic and proteomic signatures based on signatures in the human microbiome and virome as well as similar signatures from the host
- We utilize a proprietary *in vitro* model system to establish signatures and appropriate time frames early in the project, followed by *in vivo* studies to validate models
- Adopt a multi-omic approach to assess various signatures of low dose radiation exposure and employ statistical frameworks to integrate data and predict exposure states from non-invasive or trace samples

Teaming Overview and Capabilities

- SigSci has experience in human forensic analysis, microbiome analysis, genomics, proteomics, and biomarker discovery efforts following exposure to CBRNE threats
- The Contreras lab has experience in characterizing RNA-mediated regulation pathways that are affected by radiological exposures at relevant doses relevant to TEI-REX
- Institutional access to IRB, animal facilities/IACUC, and laboratories focused on genomic, proteomic, and other small molecule/metabolomic analysis
- SigSci has performed as a prime contractor on numerous IARPA programs including Proteos, FunGCAT, MAEGLIN, and multiple seedling efforts

Teaming Needs

- Open to collaborators with experience in:
 - Biodosimetry and radiation biology
 - Radiation dosimetry/health physics
 - Novel “-omics” approaches to integrate into a multi-omic approach
 - Access to relevant samples

Biophysical and biophotonic characterization of biological alterations caused by ionizing radiation

Yun Chen

Laboratory of
MEOW (Mechanical Engineering of Wet-materials)

Dept. of Mechanical Engineering

Whiting School of Engineering

Center for Cell Dynamics

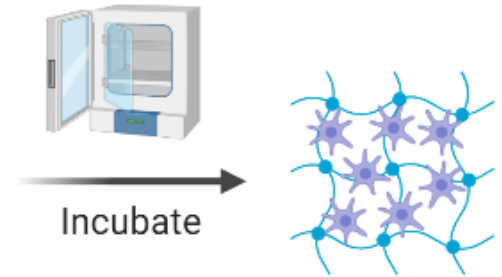
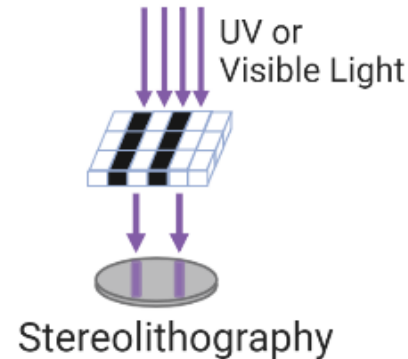
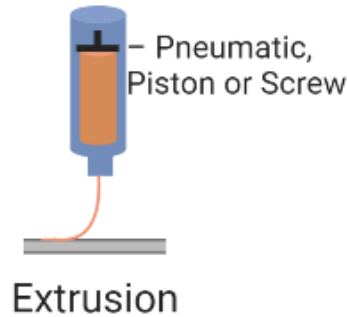
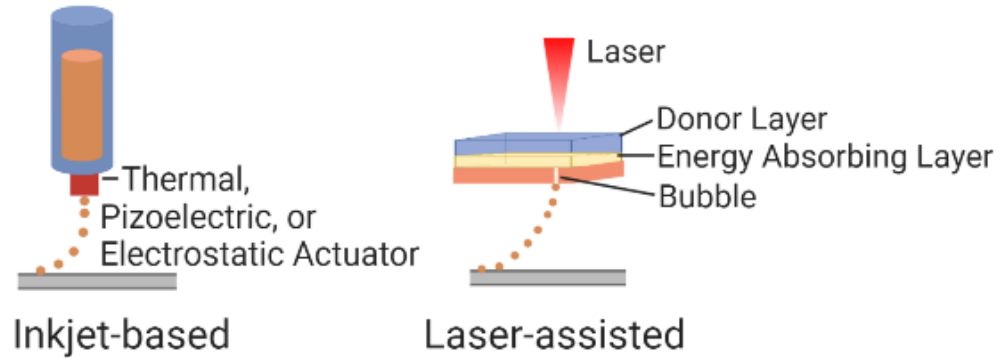
Johns Hopkins School of Medicine



Bioprinting = Placing Cells and ECM in Precision (as how they are in tissues)



Add bioink components



Culture cells in growth media

Load into bioprinter

PRE-BIOPRINTING

PROCESSING

POST-BIOPRINTING

1. Cell selection

2. Bioink selection:
mix cells with hydrogel, a water-based biomaterial

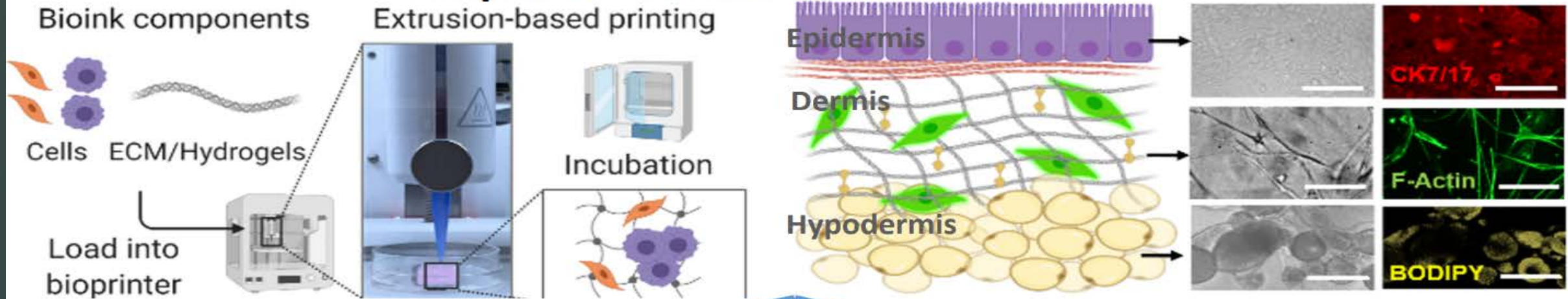
3. Morphological design:
load bioink into bioprinter print nozzle for seeding in a specific pattern, layer by layer

4. Selection of fabrication technique:
extrusion-based, inkjet-based bioprinting, laser-assisted, and stereolithography

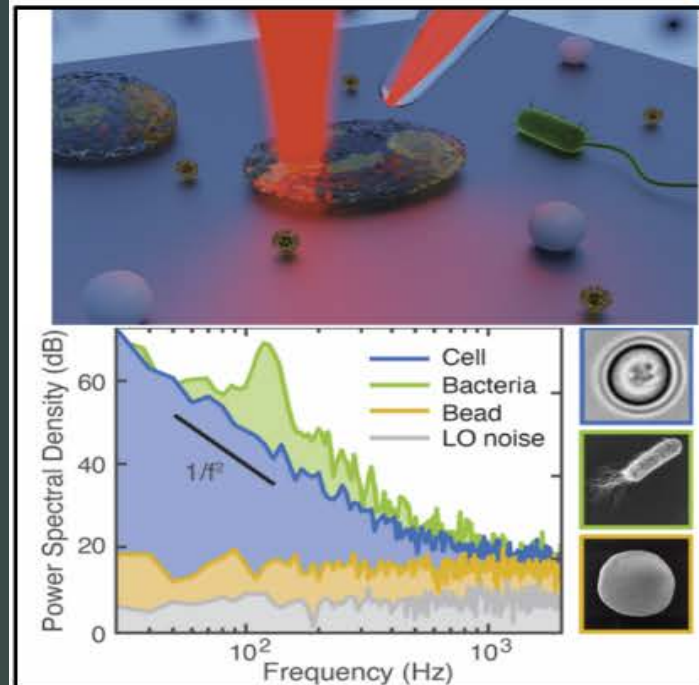
5. Stabilize bioprinted constructs with crosslinking

6. Further modifications:
removal of sacrificial inks, seed with cells

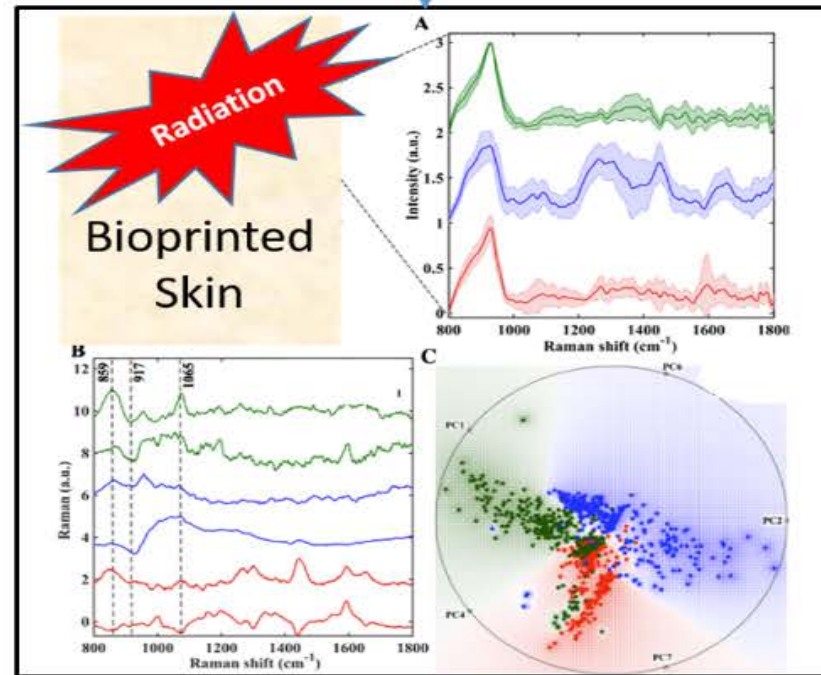
Bioprinted Skin-Mimetic Tissues



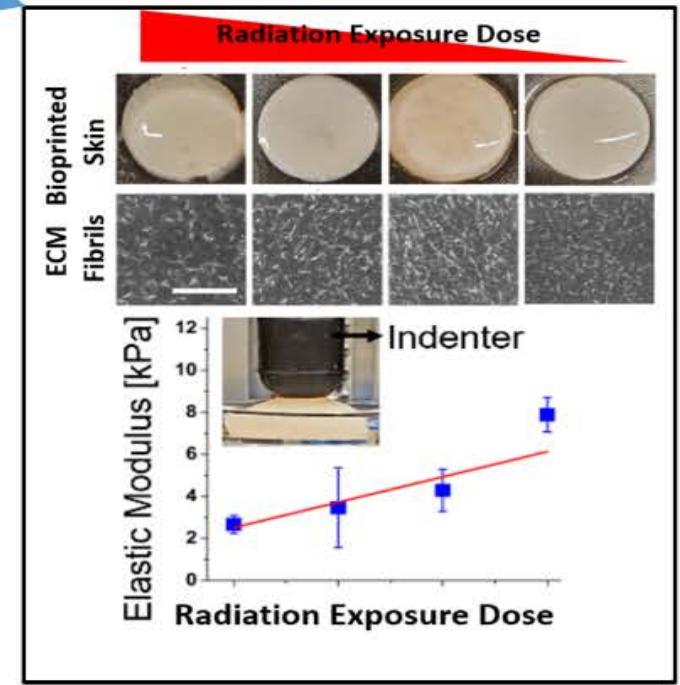
Cytoplasmic Macromolecular Motion



Raman Spectroscopy



Tissue Viscoelasticity Measurement





Turning Science Into Solutions

Advancing Biodosimetry Technologies

Sept 2021

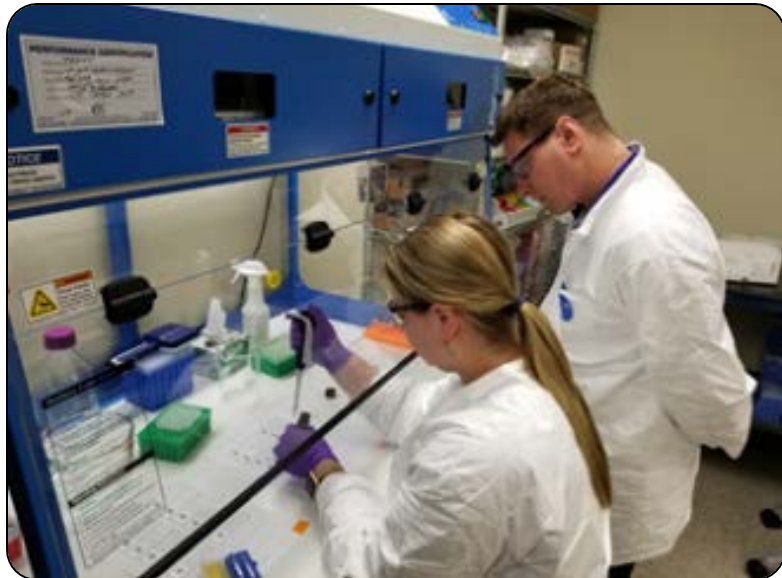
About Us

Mission → Turning Science Into Solutions



What we do: Technology development to strengthen national security and emergency preparedness

- Partnership-based model to apply emerging technologies to government priorities
- Primary focus is development of biodosimetry and R/N response technologies



Corporate Highlights

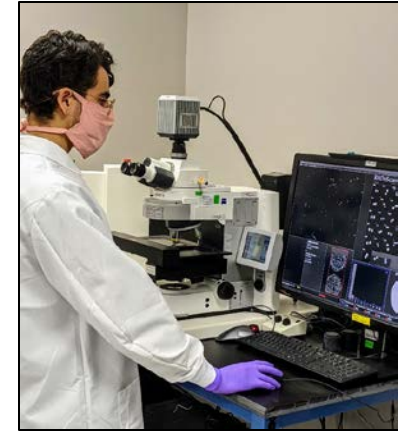
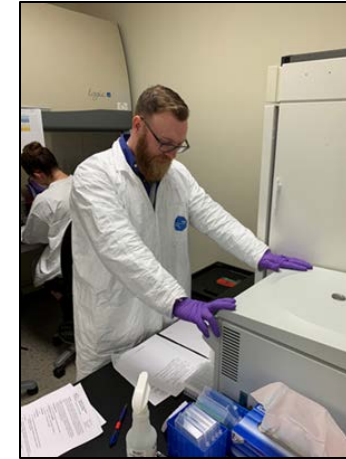
- Founded to perform advanced development of radiation diagnostics
- Team experienced in assay & instrument development, verification, validation, clinical and non-clinical research & deployment
- Extensive network of biodosimetry development partners
- Small business with established program management, contracts, and subcontracts management systems needed for USG contractor
- FDA-compliant Quality Management System
- Federal Acquisition Regulation compliant accounting system

Our People

ASELL's team brings a diverse skillset with deep biodosimetry experience



- Experienced team with wide breadth and depth of experience in technology development
- > 75% with advanced degrees
 - Molecular biology, biochemistry, chemical engineering, mathematics, biomedical engineering, systems engineering, electrical engineering
- Expertise:
 - Protein, molecular, and cell-based assay development, verification and validation
 - Algorithm development/machine learning
 - Test and evaluation
 - Ionizing radiation
 - Large and small animal models
 - Systems engineering & automation
 - Clinical research
 - Program management
 - Federal contracting
 - Regulatory affairs
 - Quality assurance



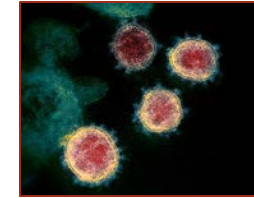
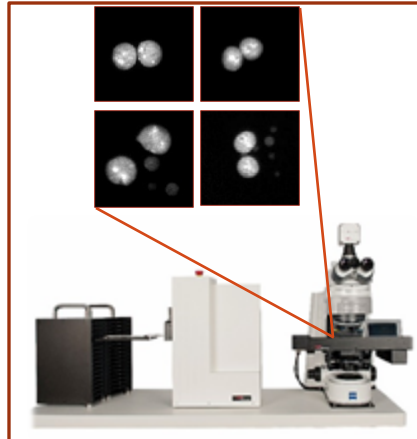
Example Dosimetry Initiatives

Meeting National Security Objectives



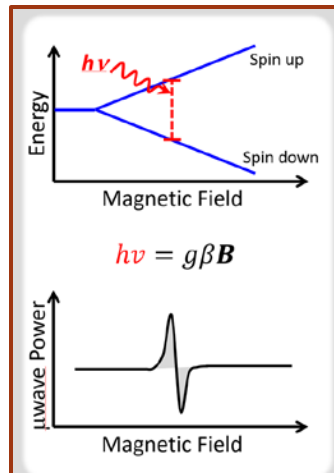
Quantitative Biodosimetry

- Quantitative, high-throughput diagnostic test (400,000 tests in 7 days) using cytokinesis-block micronucleus assay without lab-specific calibration curves
- Innovations in chemistries, cell culture formulations, image analysis and data processing algorithms
- Technology development, clinical and non-clinical studies/validation, regulatory submissions



Triage Biodosimetry

- Evaluating soluble, cell surface, and intracellular protein and hematologic biomarkers in NHP and human models
- Deployable instrument with unique imaging technology and neural network-based image processing requiring only single fingerstick sample
- Integration of instrument and biomarkers with novel biodosimetry algorithm will enable rapid, qualitative field screening for radiological exposure



Novel Electron Spin Resonance Dosimetry

- Supporting innovative spectrometer and associated technology development
- Overcoming challenges of background signatures, differentiating mix-field radiation, and rapid measurement of low dose radiation
- Leading to field-portable ESR technology for triage applications

Our Approach to TEI-REX



- Current assessment suggests the solution will likely require integration of multiple biomarkers and technologies
- Actively engaging our network of biodosimetry collaborators and evaluating emerging biomarkers and detection technologies
- Seeking interested technology partners with emerging biodosimetry solutions with promising approaches to meet specific TEI-REX objectives
- If you wish to discuss how we might collaborate, please contact Michael Ehret at mehret@asell.com

ASELL

The logo consists of the word "ASELL" in a bold, black, sans-serif font. The letter 'S' is replaced by a stylized DNA double helix. The two strands of the helix are black, and they are connected by a red horizontal bar that forms the top and bottom curves of the 'S' shape.

Turning Science Into Solutions

1



I A R P A

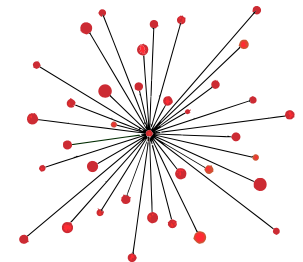
TEI-REX Proposers Day

Michael MacCoss¹, William Noble¹, Ning Cao², Eric Ford²,
Daniel Chelsky³, Paul Rudnick³

2



3



SPECTRAGEN
INFORMATICS

Proteos_UW_20210924

Team Capabilities

UW/Spectragen team addresses all key capabilities for TEI-REX program

Team Members	Affiliation	Radiation biology	In vitro and in vivo models associated with radiation exposure	Analytical biochemistry	Biomarker discovery	Biodosimetry	Machine learning and Artificial Intelligence	Radiation dosimetry/health physics	Statistics	Program management
Michael MacCoss	UW Genome Sciences			X	X					
Paul Rudnick	Spectragen Informatics			X	X					X
Daniel Chelsky	Spectragen Informatics			X	X					X
William Noble	UW Computer Sciences				X		X		X	
Ning Cao	UW Radiation Oncology	X	X	X		X		X		
Eric Ford	UW Radiation Oncology	X	X	X		X		X		

Mouse

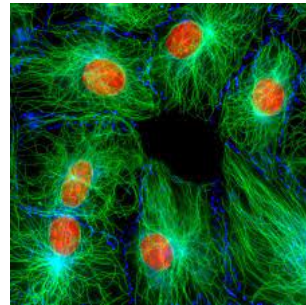
IR exposed:

Fibroblasts

- Primary dermal cells
- NIH/3T3
- L929

Skin/fur

- C57BL/6
- Nude mice



Proteomics experience with other species including rat and pig

Human

IR exposed:

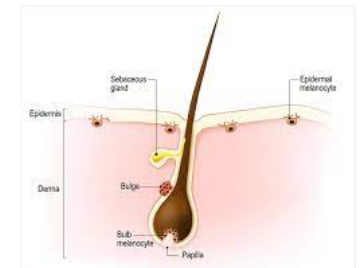
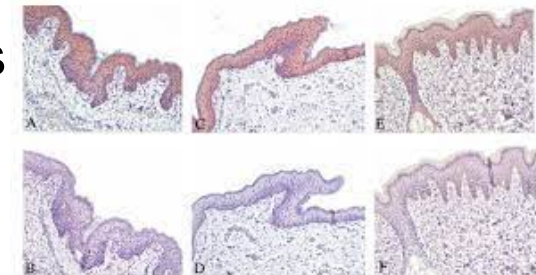
Fibroblasts

- Primary dermal cells
- Fibroblast cell lines

Human surgical skin samples

Non-IR exposed:

Human hair, hair follicles, skin adhesive disc sampling



Radiation Biology

- State of the Art Facility
- Full small animal and cell culture treatment platform
- Range of IR sources:
 - 225 kV x-rays
 - 6 MV x-rays
 - 10-50 MeV photons
 - High linear energy transfer fast neutrons



Elekta Synergy
Linear Accelerator

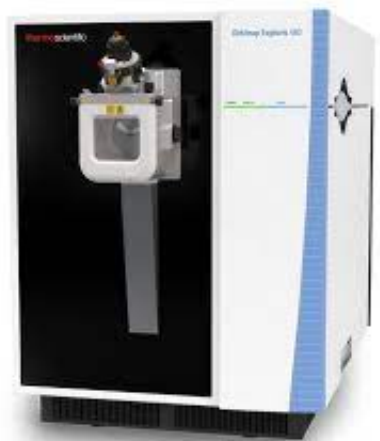


XSTRAHL small animal
radiation research platform



Scanditronix MC50 Cyclotron

Mass Spectrometry



Advanced Orbitrap instruments:

Narrow overlapping isolation window DIA

Enables comprehensive and unbiased sampling of precursor ions

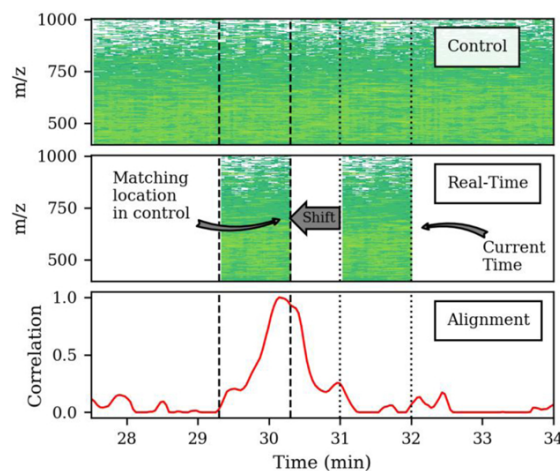
7 high performance MS instruments

Including:

Exploris 480

Fusion Lumos

Orbitrap Eclipse

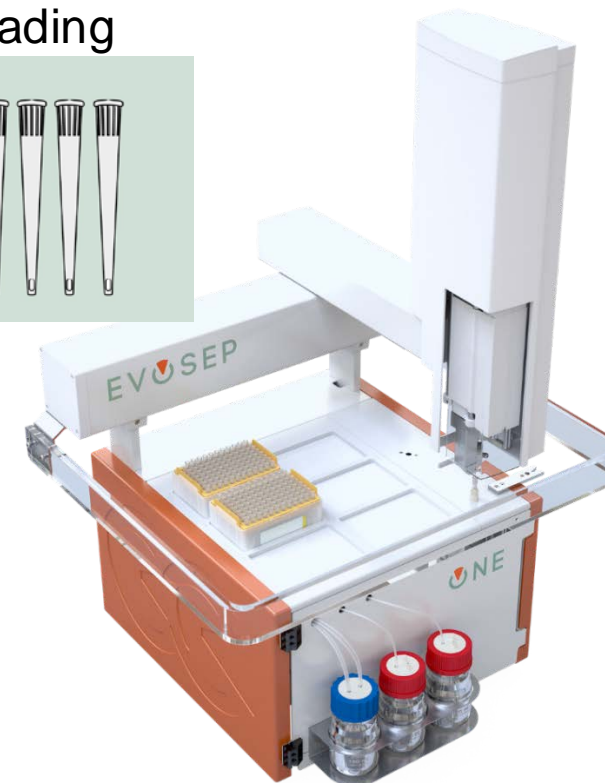
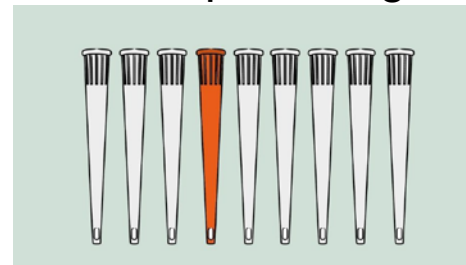


PRM: Highly multiplexed targeted proteomics

5,000 peptides quantified in 1h run

Enabled by real-time chromatographic alignment

EvoTip loading



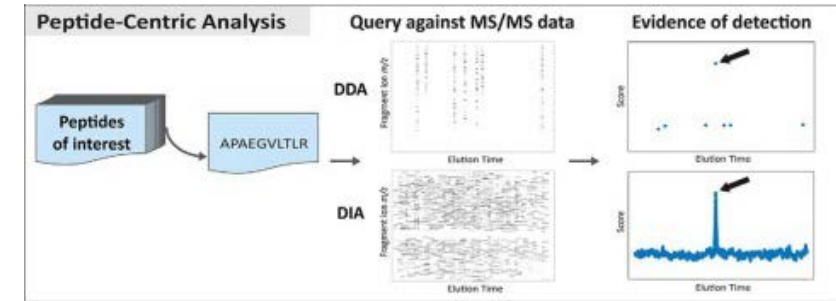
EvoSep One:

Rapid high-performance separation

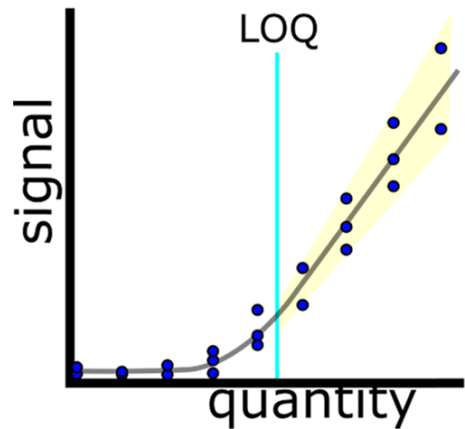
500 sample/month throughput

17-year collaboration between the Noble and MacCoss labs

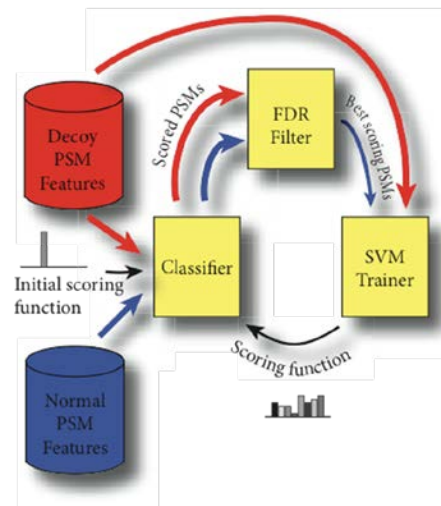
- 25 joint publications on the use of machine learning for the analysis of proteomics data
- Co-developed several of the most widely used tools
 - Percolator, Crux, Tide, accurate FDR control, etc...
 - Peptide-centric analysis for DIA
 - Analytical figures of merit, etc...



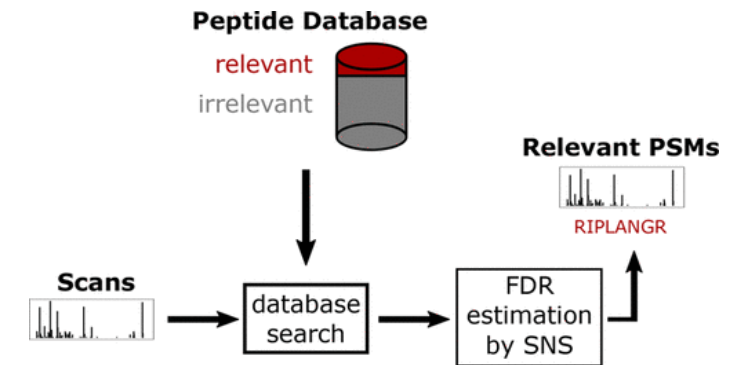
Peptide-centric proteomic analysis



Matrix-matched calibration curves for analytical figures of merit in quantitative proteomics



Percolator Algorithm



Accurately assigning peptides to spectra when only a subset of peptides are relevant